RUO

INSTRUCTION FOR USE

Pre-Plated Vaginal Women Health Panel PCR Kit

For Research Use Only

Σ 96

REF PP-VWH 005

06 2025

715 Discovery Blvd, suite 309 Cedar Park, TX 78613

Document Revision History

Rev.No_Date	Revision Description
Rev.00_June 20, 2024	First Release
Rev.01_April 10, 2025	Minor Revision
Rev.02_June 10, 2025	Minor Revision

CONTENTS

1. INTENDED USE

For Research Use Only (RUO). Not for use in diagnostic procedures. No claim or representation is intended to provide information for the diagnosis, prevention, or treatment of disease. Furthermore, this test kit is not intended for the diagnosis of infectious diseases in animals.

The ***MarinaBiolab Pre-Plated Vaginal Women Health Panel PCR Kit*** is a multiplex, qualitative Real-Time Polymerase Chain Reaction (qPCR) test intended for the simultaneous detection and identification of multiple pathogenic nucleic acids in research samples. The kit enables qPCR results in less than one hour. It is designed to detect gene sequences from the following organisms:

Targets	
Haemophilus ducreyi	*Lactobacillus gasseri*
Bacterial Vaginosis Associated Bacteria 2	*Lactobacillus iners*
Fannyhessea vaginae	*Lactobacillus crispatus*
Prevotella bivia	*Lactobacillus jensenii*
Megasphaera phylotypes 1	*Candida krusei*
Megasphaera phylotypes 2	*Candida albicans*
Mobiluncus mulieris	*Candida glabrata*
Mobiluncus curtisii	*Candida parapsilosis*
Ureaplasma urealyticum	*Candida lusitaniae*
Mycoplasma hominis	*Candida tropicalis*
Gardnerella vaginalis	*Candida dubliniensis*
Trichomonas vaginalis	*Escherichia coli*
Mycoplasma genitalium	*Streptococcus agalactiae*
Neisseria gonorrhoeae	*Staphylococcus aureus*
Chlamydia trachomatis	*Enterococcus spp*
Treponema pallidum	Herpes Simplex Virus 1
Bacteroides fragilis	Herpes Simplex Virus 2
Controls	
Human RNase P (IC)	
Bacillus atrophaeus (EC)	

2. PRINCIPLE of the PROCEDURE

DNA target regions are amplified using real-time PCR instruments, along with the specific primer and probe sets provided in the kit. During amplification, each probe binds to a specific target sequence located between the forward and reverse primers. During the extension phase of the PCR cycle, the 5' nuclease activity of Taq polymerase cleaves the probe, separating the reporter dye from the quencher and generating a fluorescent signal. With each cycle, more reporter dye molecules are released, resulting in an increase in fluorescence intensity. Fluorescence is measured at each cycle by the real-time PCR instrument. Probes labeled with distinct fluorophores are used to detect specific amplicons derived from both the target sequences and the internal control. The PCR instrument monitors the fluorescence signals in real time and interprets the data to provide a qualitative result for each target. A positive result for the presence of target DNA is indicated by the appearance of a real-time PCR amplification curve and a corresponding Cq (Quantification Cycle) value.

3. KIT COMPONENTS

The ***MarinaBiolab Pre-Plated Vaginal Women Health Panel PCR Kit*** consists of three main components:

1. qPCR Enzyme, Buffer, Forward, Reverse and Probe Mix (Pre-Plated VWHP Mix 1-11)
2. A mixture of non-infectious DNA from artificial samples, including the targets listed in the table below (PC-VWHP)
3. DNase/RNase-Free Water (NTC-VWHP)

The components of the kit are provided in Table 1-2.

Table 1. Kit components.

Component	Description	Quantity x Volume
		96 rxn PP-VWH 005
Pre-Plated VWHP Mix 1-8 **Strip 1**	Ready-to-use mix for qPCR	96 Strips (15 µL)
Pre-Plated VWHP Mix 9-11 **Strip 2**	Ready-to-use mix for qPCR	96 Strips (15 µL)
PC-VWHP	A mixture of non-infectious DNA from artificial samples, including the targets listed in the table below	2 x 400 µL
NTC- VWHP	DNase/RNase-Free Water	2 x 400 µL
PC-VWHP Pre-Mix	A mixture of non-infectious cDNA and DNA from artificial samples + Oligo + Master Mix	1 x 200 µL
NTC- VWHP Pre-Mix	DNase/RNase-Free Water + Oliog + Master Mix	1 x 200 µL

Table 2. Oligo Mix target organisms and detection channels.

Vial Name	Target	Channel
VWHP Oligo Mix 1	*Haemophilus ducreyi*	FAM
	Bacterial Vaginosis Associated Bacteria 2	HEX/VIC/JOE
	Fannyhessea vaginae	ROX/Texas Red
	Human RNase P (IC)	CY5
VWHP Oligo Mix 2	*Candida albicans*	FAM
	Candida glabrata	HEX/VIC/JOE
	Candida parapsilosis	ROX/Texas Red
	-	CY5
	Escherichia coli	FAM
	Candida dubliniensis	HEX/VIC/JOE

VWHP Oligo Mix 3	*Prevotella bivia*	ROX/Texas Red
	-	CY5

VWHP Oligo Mix 4	*Lactobacillus gasseri*	FAM
	Lactobacillus iners	HEX/VIC/JOE
	Lactobacillus crispatus	ROX/Texas Red
	-	CY5
VWHP Oligo Mix 5	*Candida lusitaniae*	FAM
	Megasphaera phylotypes 1	HEX/VIC/JOE
	Megasphaera phylotypes 2	ROX/Texas Red
	-	CY5
VWHP Oligo Mix 6	*Staphylococcus aureus*	FAM
	Mobiluncus curtisii	HEX/VIC/JOE
	Mobiluncus mulieris	ROX/Texas Red
	-	CY5
VWHP Oligo Mix 7	*Gardnerella vaginalis*	FAM
	Enterococcus spp	HEX/VIC/JOE
	Lactobacillus jensenii	ROX/Texas Red
	-	CY5
VWHP Oligo Mix 8	*Candida tropicalis*	FAM
	Trichomonas vaginalis	HEX/VIC/JOE
	Streptococcus agalactiae	ROX/Texas Red
	Bacillus atrophaeus (EC)	CY5
VWHP Oligo Mix 9	Herpes Simplex Virus 1	FAM
	Herpes Simplex Virus 2	HEX/VIC/JOE
	Mycoplasma genitalium	ROX/Texas Red
	-	CY5
VWHP Oligo Mix 10	*Neisseria gonorrhoeae*	FAM
	Candida krusei	HEX/VIC/JOE
	Chlamydia trachomatis	ROX/Texas Red
	-	CY5
VWHP Oligo Mix 11	*Treponema pallidum*	FAM
	Ureaplasma urealyticum	HEX/VIC/JOE
	Mycoplasma hominis	ROX/Texas Red
	Bacteroides fragilis	CY5

The oligonucleotide set targeting the human *RNase P* (Internal Control: IC) and *Bacillus atrophaeus* (External Control: EC) are used to monitor sampling, nucleic acid extraction, and inhibition of qPCR. The kit also contains negative and positive control templates to evaluate contamination and the qPCR reagent stability, respectively.

4. EQUIPMENT and MATERIALS REQUIRED but NOT PROVIDED

- 2-8°C Refrigerator
- ≤ -20°C Freezer
- ≤ -70°C Freezer (Optional)
- Vortex mixer
- Benchtop centrifuge with rotor for 1.5 mL tubes
- Benchtop mini centrifuge with rotor for PCR strips
- Benchtop plate centrifuge
- Biological Safety Cabinet (BSC)
- PCR cabinet for PCR Setup
- Adjustable Micropipettes: 1-10, 10-100, 100-1000 µL
- Sterile DNase/RNase free micropipettes tips - Compatible with the micropipettes
- Cold tube rack for microfuge tubes (1.5/2 mL) and for PCR tubes (0.1/0.2 mL)
- Disposable, powder-free, nitrile gloves
- Disposable (preferably) laboratory coat
- Surface decontaminants - Freshly diluted 10% bleach solution (0.5% NaClO)
- Applied Biosystems QuantStudio 5, 7, and 12K with Design & Analysis software and consumables
- Bio-Rad CFX96 Touch™/CFX96™ Dx/CFX Opus 96™/CFX Opus 96™ Dx with Maestro software v1.1 and consumables

5. WARNING and PRECAUTIONS

- The ***MarinaBiolab Pre-Plated Vaginal Women Health Panel PCR Kit*** is intended for research use only and should be used by professionally trained, qualified personnel. All procedures should be performed in accordance with Good Laboratory Practices (GLP).
- Biological material used for nucleic acid extraction should be handled as potentially infectious. Appropriate safety precautions are recommended when handling biological material (e.g., do not pipet by mouth; wear disposable gloves; disinfect hands after completing the test).
- Biological material should be inactivated before disposal (e.g., autoclaving). Disposable items should be autoclaved or incinerated after use.
- In the event of a spill involving potentially infectious materials, the spill should be immediately absorbed with paper tissue, and the affected area should be disinfected using a suitable standard disinfectant or 70% alcohol. Materials used for cleaning spills, including gloves, should be inactivated before disposal (e.g., autoclaving).
- Disposal of all samples, unused reagents and waste should be in accordance with country, federal, state, and local regulations.
- To avoid microbial contamination of reagents during aliquoting, it is recommended to use sterile, single-use pipettes and tips. Reagents that appear cloudy or show signs of microbial contamination should not be used.
- The kit should be stored away from nucleic acid sources and PCR amplicons to prevent contamination.
- Always check the expiration date on the kit. Do not use expired or improperly stored kits.
- Components in the kit should not be mixed with components from different lot numbers or from different manufacturers, even if they contain the same components.
- The kit components should be gently mixed before use by shaking.
- A common issue with PCR-based assays is false positive results caused by contamination from PCR amplicons. To minimize the risk of amplicon contamination:
    - Ensure separate work areas with dedicated apparatus are available for each stage of the procedure.
    - Do not open reaction tubes/plates post-amplification to avoid contamination with amplicons.
    - Discard used tubes/plates immediately in a biohazard container after completing the run.
    - Minimize handling of tubes/plates after testing.
    - Change gloves after handling used tubes/plates.

6. HANDLING, STORAGE, and STABILITY

- The ***MarinaBiolab Pre-Plated Vaginal Women Health Panel PCR Kit*** is shipped on dry ice. If any component is not frozen upon arrival or if the outer packaging has been compromised during shipment, please contact **MarinaBiolab** or the local distributor immediately.
- Upon arrival, all components should be stored between -25°C and -15°C.
- Repeated freezing and thawing of the kit components may reduce detection quality. The kit can withstand up to 15 freeze/thaw cycles without impacting performance.
- When stored under the specified conditions, the kit remains stable until the expiration date printed on the package. The expiration date is 12 months from the date of manufacture.
- All components must be thawed at ambient temperature for at least 30 minutes before use.
- It is recommended to keep all components on ice when preparing the assay mixes.
- The primer and probe mixes contain fluorophore-labeled probes and should be protected from direct sunlight and prolonged exposure to ambient light.
- Do not use expired or improperly stored components.

7. TEST PROCEDURE

7.1. Sample Preparation and Nucleic Acid Extraction

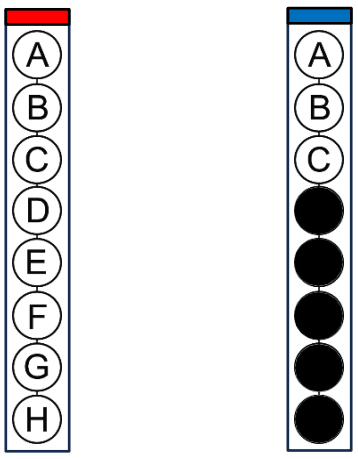
Samples intended for nucleic acid isolation must be collected using appropriate cell collection systems. The performance of the kit is highly dependent on both the quantity and quality of the extracted nucleic acid. Ensure that the extraction method used is compatible with real-time PCR technology.

If the laboratory's established standard protocol is used for nucleic acid isolation, it must be validated by the end user.

For frozen samples or previously extracted nucleic acid, thaw only the amount required for testing on the same day. Avoid multiple freeze/thaw cycles, as these can compromise nucleic acid integrity. For best results, use the nucleic acid immediately after thawing.

7.2. PCR Reaction Preparation and Processing

- Determine the number of reactions needed and prepare a PCR plate layout accordingly.
- The plate layout should include the following:
    - Reactions for each test sample and extraction negative control.
    - PCR control reactions:
        - Positive Control (provided in the kit)
        - Negative (No Template) Control (NTC) (provided in the kit)
- Completely thaw all components at room temperature for at least 30 minutes prior to use.
- When they thaw, vortex and **spin down** briefly the components and place them on cold block during the whole test procedure.
- Use 2 strips (Strip 1 and Strip 2) for each sample or control.
- The orientations of Strip 1 and 2 should be as shown below.



Strip 1 **Strip 2**

- Open carefully the strips (gently open it from the side way, DO NOT PEEL OFF THE LID) and add 5 µL of the isolated sample or control to the corresponding wells or 20 µL of Pre-Mix PC or NTC to an empty well.
- The final reaction mix volume is 20 µL.
- Re-cap the strips and **spin down** for 5 seconds.
- Insert strips into the real-time PCR instrument and amplify according to the following PCR profile.

For each run, use one well of PC-Mix and one well of NTC-Mix as shown in the diagram below.

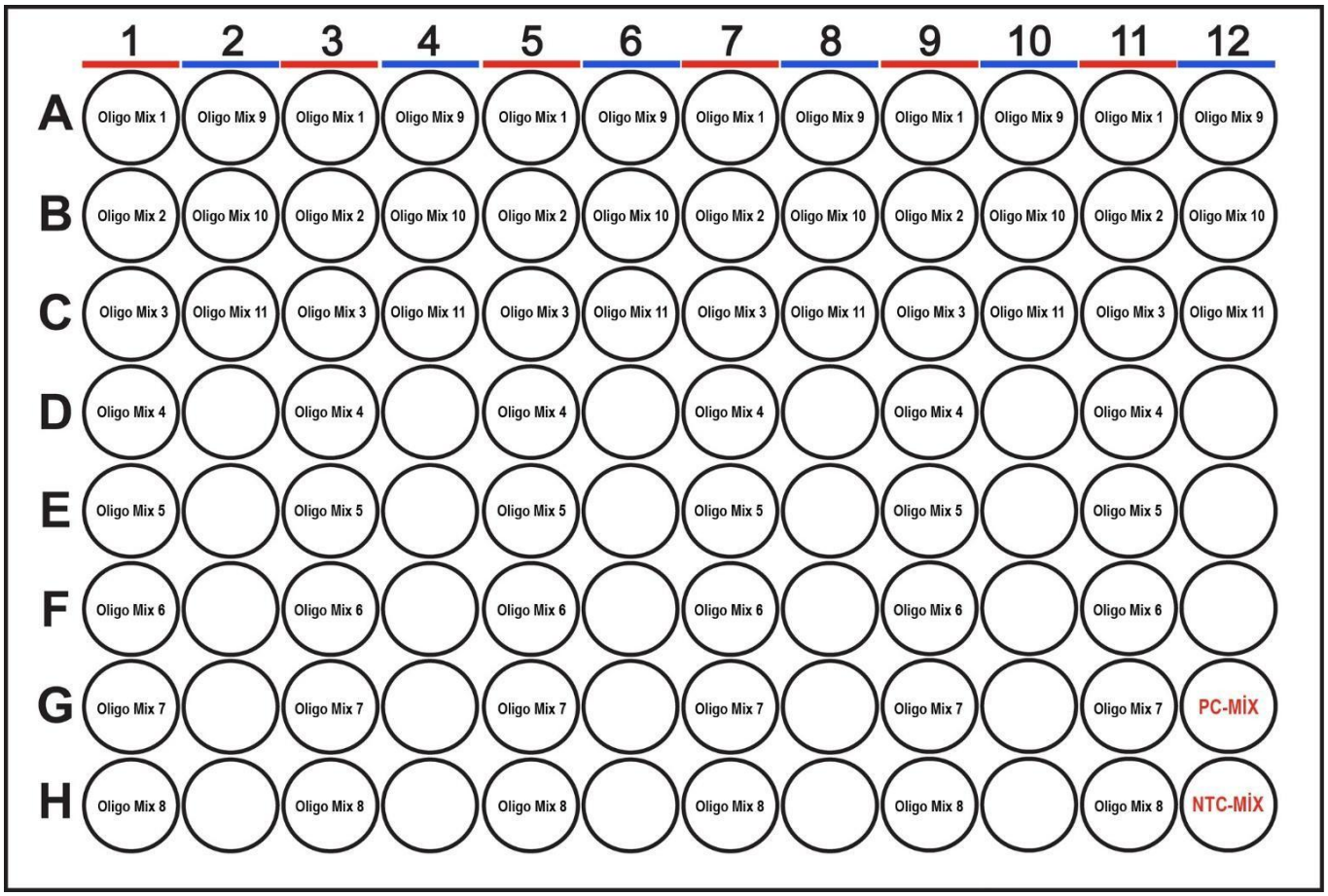


Table 3. Amplification profile.

Step	Number of Cycles	Temperature	Time	Data Collection
Initial Denaturation	1	95 °C	10 sec	FAM
Denaturation	40	95 °C	5 sec	HEX/VIC/JOE
Annealing/Extension		55 °C	15 sec	ROX/Texas Red CY5

8. INTERPRETATION OF RESULTS

MarinaBiolab Pre-Plated Vaginal Women Health Panel PCR Kit provides a qualitative result for the presence (Detected) or absence (Not Detected) of the target genes.

8.1. Calculation of Cq Values and Instrument-Specific Requirements

Configure the following instrument settings before evaluating the results.

Table 4. Instrument-specific settings.

Instrument	Threshold Level	Other Settings
CFX96 Touch™/CFX96™ Dx/CFX Opus 96™/CFX Opus 96™ Dx (Bio-Rad)	500 RFU	-
QuantStudio™ 5, 7 and 12K (Applied Biosystems™)	Auto	-

The shape of the amplification curves should be evaluated. If the instrument's software assigns a Cq value to a sample and the curve is sigmoidal, the Cq value can be used in the final assessment. *Non-sigmoidal curves should be recorded as negative.*

A result is considered positive if the Cq value is ≤35, or as determined by your laboratory's protocols.

8.2. Overall Validity of Detection

Table 5. Expected performance of controls.

Control Type	Used to Monitor	Signal	
		Target Channel	Internal/External Control Channel
Negative Control	Cross-contamination during extraction and reaction setup	-	-
No template addition	Reagent and/or environmental contamination	-	-
Positive Control	qPCR reaction setup and reagent integrity	+	+
Internal/External Control	To monitor the integrity of nucleic acid extraction and qPCR from each specimen	Not applicable	+

Before analyzing sample results, we recommend verifying the validity of the real-time PCR test. For each run, please confirm that the Positive and Negative controls performed as expected, based on the following criteria:

Table 6. Run validity/positive and negative control pass criteria.

Positive Control		Negative Control		Results	Recommendation
Target Channel	Internal/External Control Channel	Target Channel	Internal/External Control Channel		
+	+	-	-	VALID	Proceed with the interpretation of sample results.

Any of them is Negative	Not considered	INVALID	Contact the manufacturer, replenish the reagents, and repeat the reaction.
Not considered	Any of them is Positive	INVALID	Repeat the analysis, ensuring to follow the 'Warnings and Precautions' outlined in the IFU.

If any control fails to perform as described above, the run is considered invalid and must be repeated. If the issue persists, contact the manufacturer.

If all controls perform as expected, proceed with the interpretation of the results.

8.3. Interpretation of Unknown Specimen Results

The data generated by the instruments can be manually evaluated and reported using their software.

Table 7. Interpretation of unknown specimen results for DNA pathogens.

DNA Pathogens	Internal Control (*RNase P*)	External Control (*Bacillus atrophaeus*)	Results	Interpretation
Positive (+) (Cq<35)	Positive (+) (Cq<35)	Positive (+) (Cq<35)	Positive for Target	Target DNA is detected
Positive (+) (Cq<35)	Negative (-) (Cq≥35 or N/A)	Positive (+) (Cq<35)	Positive for Target	Target DNA is detected
Positive (+) (Cq<35)	Positive (+) (Cq<35)	Negative (-) (Cq≥35 or N/A)	Positive for Target	Target DNA is detected
Positive (+) (Cq<35)	Negative (-) (Cq≥35 or N/A)	Negative (-) (Cq≥35 or N/A)	Invalid	Repeat the test by re-extracting the sample. If the result remains invalid, consider collecting a new sample.
Negative (-) (Cq≥35 or N/A)	Positive (+) (Cq<35)	Positive (+) (Cq<35)	Negative for Target	Target DNA is not detected
Negative (-) (Cq≥35 or N/A)	Negative (-) (Cq≥35 or N/A)	Positive (+) (Cq<35)	Negative for Target	Target DNA is not detected
Negative (-) (Cq≥35 or N/A)	Positive (+) (Cq<35)	Negative (-) (Cq≥35 or N/A)	Negative for Target	Target DNA is not detected
Negative (-) (Cq≥35 or N/A)	Negative (-) (Cq≥35 or N/A)	Negative (-) (Cq≥35 or N/A)	Invalid	Repeat the test by re-extracting the sample. If the result remains invalid, consider collecting a new sample.

9. ASSAY LIMITATIONS

- The ***MarinaBiolab Pre-Plated Vaginal Women Health Panel PCR Kit*** is intended for use only by professionally trained and qualified staff.
- A false negative result may occur if the specimen is improperly collected, transported, or handled. False negatives can also occur if amplification inhibitors are present in the specimen or if insufficient numbers of organisms are present.
- Spontaneous mutations within the target sequences may result in failure to detect the target. While the test design mitigates this risk, if target detection failure is anticipated, it is recommended to test the specimen with a different assay that targets other sequences in the genome.
- There is a risk of false positive results due to cross-contamination by target viruses and/or bacteria, their nucleic acids or amplified products, or from non-specific signals in the assay. Proper handling of consumables, as outlined in the Warnings and Precautions section, is crucial to minimize this risk.
- This assay is qualitative and does not provide a quantitative assessment of the detected organism's concentration.
- All instruments (e.g., pipettes, real-time PCR cyclers) must be calibrated according to the manufacturer's instructions.

10. PERFORMANCE CHARACTERISTICS

10.1. Analytical Sensitivity (Limit of Detection, LoD)

The limit of detection (LoD) was defined as the concentration at which the test produces a positive result more than 95% of the time. Serial dilutions of the strains were tested, and the initial tentative LoD was confirmed with twenty (20) replicates. To ensure the accuracy of the LoD determination, if the initial detection rate was 100%, an additional twenty (20) replicates were performed at the next lower concentration until a detection rate of ≤95% was achieved.

For nucleic acid extraction, a simulated research matrix was spiked with strains and processed using the Automatic Nucleic Acids Extraction Instrument. Testing was carried out on the CFX96 Touch™ (Bio-Rad) Real-Time PCR system. The confirmed LoDs for the strains tested, along with the corresponding LoDs for the ***MarinaBiolab Pre-Plated Vaginal Women Health Panel PCR Kit*** reportable targets, are presented in Table 8 below.

Table 8. Summary of LoD study results.

Analyte	Isolate ID/Source	LoD Concentration (copies/mL)	Detected/Total
Haemophilus ducreyi	Zeptometrix 0801736DNA	2.5E+01 copies/mL	**20/20** 100%
Bacterial Vaginosis Associated Bacteria 2	ATCC TSD-275	6.7+01 copies/mL	**20/20** 100%
Fannyhessea vaginae	ATCC BAA-55	7.8+01 copies/mL	**20/20** 100%
Candida albicans	ATCC 10231	3.4E+02 copies/mL	**20/20** 100%
Candida glabrata	ATCC 90030	4.4E+01 copies/mL	**20/20** 100%
Candida parapsilosis	ATCC 22019	5.8E+01 copies/mL	**20/20** 100%
Escherichia coli	ATCC 25922	3.5E+01 copies/mL	**20/20** 100%
Streptococcus agalactiae	ATCC 12386	6.7E+01 copies/mL	**19/20** 95%
Prevotella bivia	Zeptometrix 0801756	8.9+01 copies/mL	**20/20** 100%
Lactobacillus gasseri	Zeptometrix 0804327	1.1+02 copies/mL	**20/20** 100%
Lactobacillus iners	Zeptometrix 0804261	7.7+01 copies/mL	**20/20** 100%
Lactobacillus crispatus	Zeptometrix 0804143	9.5+01 copies/mL	**20/20** 100%

Candida lusitaniae	Zeptometrix 0801603	8.0+01 copies/mL	**20/20** 100%
Megasphaera phylotypes 1	In-house	2.1+02 copies/mL	**20/20** 100%
Megasphaera phylotypes 2	In-house	1.9+02 copies/mL	**20/20** 100%
Staphylococcus aureus	ATCC 12600	8.6E+01 copies/mL	**20/20** 100%
Mobiluncus curtisii	Zeptometrix 0804141	8.9+01 copies/mL	**20/20** 100%
Mobiluncus mulieris	Zeptometrix 0804116	9.1+01 copies/mL	**20/20** 100%
Gardnerella vaginalis	ATCC 49145	1.5E+01 copies/mL	**20/20** 100%
Enterococcus faecalis	Zeptometrix 0804216	3.6E+02 copies/mL	**20/20** 100%
Enterococcus faecium	ATCC BAA-2127	4.5E+01 copies/mL	**20/20** 100%
Lactobacillus jensenii	Zeptometrix 0804260	8.5+01 copies/mL	**20/20** 100%
Candida tropicalis	ATCC 750	5.7E+01 copies/mL	**20/20** 100%
Trichomonas vaginalis	ATCC 30001	2.5E+01 copies/mL	**20/20** 100%
Herpes Simplex Virus 1	ATCC VR-1778	1.2E+02 copies/mL	**20/20** 100%
Herpes Simplex Virus 2	Zeptometrix 0810217CF	3.7E+01 copies/mL	**20/20** 100%
Mycoplasma genitalium	ATCC 33530D	4.0E+01 copies/mL	**19/20** 95%
Candida dubliniensis	Zeptometrix 0801915	7.8E+01 copies/mL	**20/20** 100%
Neisseria gonorrhoeae	ATCC 19424	6.0E+01 copies/mL	**20/20** 100%
Candida krusei	ATCC 2159	6.8E+01 copies/mL	**20/20** 100%
Chlamydia trachomatis	Zeptometrix 0801775	5.8E+01 copies/mL	**20/20** 100%

Treponema pallidum	In-house	6.9E+01 copies/mL	**20/20** 100%
Ureaplasma urealyticum	ATCC 27618	1.1E+02 copies/mL	**20/20** 100%
Mycoplasma hominis	ATCC 27545-TTR	1.1E+02 copies/mL	**20/20** 100%
Bacteroides fragilis	Zeptometrix 0801583	6.9E+01 copies/mL	**20/20** 100%

10.2. Device Equivalence Study

A device equivalence study was conducted to assess the differences in results obtained using the kit across various instruments. For this purpose, the same LoD determination study was repeated using the Bio-Rad CFX96™ Dx/CFX Opus 96™/CFX Opus 96™ Dx/ CFX384 Touch™/CFX Opus 384™, Applied Biosystems QuantStudio 5, 7, and 12K, Qiagen Rotor-Gene Q 5plex Platform, and Roche LightCycler 480. Similar results were obtained at the 1x LoD concentration level of the targets in the device equivalence study across the different instruments.

10.3. Analytical Reactivity (Inclusivity)

10.3.1. In-Slico Analytical Reactivity

A BLAST search of the oligonucleotides was conducted on the genome sequences of *Haemophilus ducreyi, Bacterial Vaginosis Associated Bacteria 2, Fannyhessea vaginae, Candida albicans, Candida glabrata, Candida parapsilosis, Escherichia coli, Streptococcus agalactiae, Prevotella bivia, Lactobacillus gasseri, Lactobacillus iners, Lactobacillus crispatus, Candida lusitaniae, Megasphaera phylotypes 1, Megasphaera phylotypes 2, Staphylococcus aureus, Mobiluncus curtisii, Mobiluncus mulieris, Gardnerella vaginalis, Enterococcus spp, Lactobacillus jensenii, Candida tropicalis, Trichomonas vaginalis*, Herpes Simplex Virus 1, Herpes Simplex Virus 2, *Mycoplasma genitalium, Candida dubliniensis, Neisseria gonorrhoeae, Candida krusei, Chlamydia trachomatis, Treponema pallidum, Ureaplasma urealyticum, Mycoplasma hominis,* and *Bacteroides fragilis* using the Primer-BLAST tool on the NCBI database.

The aggregated results of all in-silico analyses performed using the NCBI database are provided in the table below. The melting temperatures (Tm) of the oligonucleotide sequences with a 1-base mismatch remain higher than the annealing temperature specified in the PCR cycle parameters of the kit. Therefore, single base mismatches in the sequences are not expected to impact the inclusivity of the test.

Table 9. In-silico analysis results performed in the NCBI database.

Target	Primer	Total number of target sequences	Ratio of the sequences without mismatch	Ratio of the sequences with 1 base mismatch	Ratio of the sequences with 2 base mismatches	Ratio of the sequences with 3 base mismatches
Haemophilus ducreyi	Sense Primer	40	100.00%	0.00%	0.00%	0.00%
Haemophilus ducreyi	Antisense Primer	40	100.00%	0.00%	0.00%	0.00%
Haemophilus ducreyi	Hydrolysis Probe	40	100.00%	0.00%	0.00%	0.00%

Bacterial Vaginosis Associated Bacteria 2	Sense Primer	24	100.00%	0.00%	0.00%	0.00%
Bacterial Vaginosis Associated Bacteria 2	Antisense Primer	24	100.00%	0.00%	0.00%	0.00%
Bacterial Vaginosis Associated Bacteria 2	Hydrolysis Probe	24	100.00%	0.00%	0.00%	0.00%
Fannyhessea vaginae	Sense Primer	44	100.00%	0.00%	0.00%	0.00%
Fannyhessea vaginae	Antisense Primer	44	100.00%	0.00%	0.00%	0.00%
Fannyhessea vaginae	Hydrolysis Probe	44	100.00%	0.00%	0.00%	0.00%
Candida albicans	Sense Primer	3.629	99.69%	0.31%	0.00%	0.00%
Candida albicans	Antisense Primer	3.728	98.85%	2.25%	0.00%	0.00%
Candida albicans	Hydrolysis Probe	3.728	98.52%	2.48%	0.00%	0.00%
Candida glabrata	Sense Primer	763	100%	0.00%	0.00%	0.00%
Candida glabrata	Antisense Primer	1.111	99.20%	0.80%	0.00%	0.00%
Candida glabrata	Hydrolysis Probe	1.111	99.64%	0.36%	0.00%	0.00%
Candida parapsilosis	Sense Primer	2.559	99.74%	0.26%	0.00%	0.00%
Candida parapsilosis	Antisense Primer	2.463	100%	0.00%	0.00%	0.00%
Candida parapsilosis	Hydrolysis Probe	2.463	100%	0.00%	0.00%	0.00%
Escherichia coli	Sense Primer	5.547	99.25%	0.75%	0.00%	0.00%
Escherichia coli	Antisense Primer	5.579	99.65%	0.35%	0.00%	0.00%
Escherichia coli	Hydrolysis Probe	5.579	99.78%	0.22%	0.00%	0.00%
Streptococcus agalactiae	Sense Primer	226	99.95%	0.05%	0.00%	0.00%
Streptococcus agalactiae	Antisense Primer	236	100.00%	0.00%	0.00%	0.00%
Streptococcus agalactiae	Hydrolysis Probe	236	100.00%	0.00%	0.00%	0.00%
Prevotella bivia	Sense Primer	56	99.80%	0.20%	0.00%	0.00%
Prevotella bivia	Antisense Primer	56	99.80%	0.20%	0.00%	0.00%
Prevotella bivia	Hydrolysis Probe	52	99.60%	0.40%	0.00%	0.00%
Lactobacillus gasseri	Sense Primer	987	98.24%	1.76%	0.00%	0.00%
Lactobacillus gasseri	Antisense Primer	987	98.24%	1.76%	0.00%	0.00%
Lactobacillus gasseri	Hydrolysis Probe	956	98.10%	1.90%	0.00%	0.00%
Lactobacillus iners	Sense Primer	1458	97.68%	2.00%	0.32%	0.00%
Lactobacillus iners	Antisense Primer	1458	97.68%	2.00%	0.32%	0.00%
Lactobacillus iners	Hydrolysis Probe	1402	97.70%	2.20%	0.10%	0.00%
Lactobacillus crispatus	Sense Primer	825	98.21%	1.79%	0.00%	0.00%

Lactobacillus crispatus	Antisense Primer	825	98.21%	1.79%	0.00%	0.00%
Lactobacillus crispatus	Hydrolysis Probe	828	98.11%	1.89%	0.00%	0.00%
Candida lusitaniae	Sense Primer	220	99.60%	0.40%	0.00%	0.00%
Candida lusitaniae	Antisense Primer	220	99.60%	0.40%	0.00%	0.00%
Candida lusitaniae	Hydrolysis Probe	220	99.80%	0.20%	0.00%	0.00%
Megasphaera phylotypes 1	Sense Primer	54	99.80%	0.20%	0.00%	0.00%
Megasphaera phylotypes 1	Antisense Primer	54	99.80%	0.20%	0.00%	0.00%
Megasphaera phylotypes 1	Hydrolysis Probe	53	99.60%	0.40%	0.00%	0.00%
Megasphaera phylotypes 2	Sense Primer	20	100.00%	0.00%	0.00%	0.00%
Megasphaera phylotypes 2	Antisense Primer	20	100.00%	0.00%	0.00%	0.00%
Megasphaera phylotypes 2	Hydrolysis Probe	20	100.00%	0.00%	0.00%	0.00%
Staphylococcus aureus	Sense Primer	657	99.80%	0.20%	0.00%	0.00%
Staphylococcus aureus	Antisense Primer	657	99.80%	0.20%	0.00%	0.00%
Staphylococcus aureus	Hydrolysis Probe	655	99.70%	0.30%	0.00%	0.00%
Mobiluncus curtisii	Sense Primer	34	100.00%	0.00%	0.00%	0.00%
Mobiluncus curtisii	Antisense Primer	34	100.00%	0.00%	0.00%	0.00%
Mobiluncus curtisii	Hydrolysis Probe	34	100.00%	0.00%	0.00%	0.00%
Mobiluncus mulieris	Sense Primer	20	100.00%	0.00%	0.00%	0.00%
Mobiluncus mulieris	Antisense Primer	20	100.00%	0.00%	0.00%	0.00%
Mobiluncus mulieris	Hydrolysis Probe	20	100.00%	0.00%	0.00%	0.00%
Gardnerella vaginalis	Sense Primer	52	100.00%	0.00%	0.00%	0.00%
Gardnerella vaginalis	Antisense Primer	52	100.00%	0.00%	0.00%	0.00%
Gardnerella vaginalis	Hydrolysis Probe	50	100.00%	0.00%	0.00%	0.00%
Enterococcus spp	Sense Primer	1652	100.00%	0.00%	0.00%	0.00%
Enterococcus spp	Antisense Primer	1652	100.00%	0.00%	0.00%	0.00%
Enterococcus spp	Hydrolysis Probe	1640	99.89%	0.11%	0.00%	0.00%
Lactobacillus jensenii	Sense Primer	687	98.23%	2.67%	0.00%	0.00%
Lactobacillus jensenii	Antisense Primer	687	98.23%	2.67%	0.00%	0.00%
Lactobacillus jensenii	Hydrolysis Probe	660	98.07%	2.93%	0.00%	0.00%
Candida tropicalis	Sense Primer	1.164	98.40%	2.60%	0.00%	0.00%
Candida tropicalis	Antisense Primer	1.906	97.83%	2.17%	0.00%	0.00%
Candida tropicalis	Hydrolysis Probe	1.906	97.12%	2.88%	0.00%	0.00%

Trichomonas vaginalis	Sense Primer	63	99.79%	0.21%	0.00%	0.00%
Trichomonas vaginalis	Antisense Primer	63	99.79%	0.21%	0.00%	0.00%
Trichomonas vaginalis	Hydrolysis Probe	60	99.75%	0.25%	0.00%	0.00%
Herpes Simplex Virus 1	Sense Primer	622	99.67%	0.23%	0.00%	0.00%
Herpes Simplex Virus 1	Antisense Primer	625	99.25%	0.75%	0.00%	0.00%
Herpes Simplex Virus 1	Hydrolysis Probe	625	99.20%	0.80%	0.00%	0.00%
Herpes Simplex Virus 2	Sense Primer	454	100%	0.00%	0.00%	0.00%
Herpes Simplex Virus 2	Antisense Primer	462	99.81%	0.19%	0.00%	0.00%
Herpes Simplex Virus 2	Hydrolysis Probe	462	99.89%	0.11%	0.00%	0.00%
Mycoplasma genitalium	Sense Primer	50	100.00%	0.00%	0.00%	0.00%
Mycoplasma genitalium	Antisense Primer	50	100.00%	0.00%	0.00%	0.00%
Mycoplasma genitalium	Hydrolysis Probe	48	100.00%	0.00%	0.00%	0.00%
Candida dubliniensis	Sense Primer	452	96.44%	3.56%	0.00%	0.00%
Candida dubliniensis	Antisense Primer	452	96.44%	3.56%	0.00%	0.00%
Candida dubliniensis	Hydrolysis Probe	420	98.22%	1.78%	0.00%	0.00%
Neisseria gonorrhoeae	Sense Primer	597	99.20%	0.80%	0.00%	0.00%
Neisseria gonorrhoeae	Antisense Primer	597	99.20%	0.80%	0.00%	0.00%
Neisseria gonorrhoeae	Hydrolysis Probe	590	99.05%	0.80%	0.05%	0.00%
Candida krusei	Sense Primer	1.415	100%	0.00%	0.00%	0.00%
Candida krusei	Antisense Primer	1.415	100%	0.00%	0.00%	0.00%
Candida krusei	Hydrolysis Probe	1.415	100%	0.00%	0.00%	0.00%
Chlamydia trachomatis	Sense Primer	862	99.64%	0.36%	0.00%	0.00%
Chlamydia trachomatis	Antisense Primer	862	99.64%	0.36%	0.00%	0.00%
Chlamydia trachomatis	Hydrolysis Probe	846	99.60%	0.40%	0.00%	0.00%
Treponema pallidum	Sense Primer	538	99.64%	0.36%	0.00%	0.00%
Treponema pallidum	Antisense Primer	538	99.64%	0.36%	0.00%	0.00%
Treponema pallidum	Hydrolysis Probe	538	99.50%	0.50%	0.00%	0.00%
Ureaplasma urealyticum	Sense Primer	90	99.90%	0.10%	0.00%	0.00%
Ureaplasma urealyticum	Antisense Primer	90	99.90%	0.10%	0.00%	0.00%
Ureaplasma urealyticum	Hydrolysis Probe	88	99.90%	0.10%	0.00%	0.00%
Mycoplasma hominis	Sense Primer	48	100.00%	0.00%	0.00%	0.00%
Mycoplasma hominis	Antisense Primer	48	100.00%	0.00%	0.00%	0.00%

Mycoplasma hominis	Hydrolysis Probe	48	100.00%	0.00%	0.00%	0.00%
Bacteroides fragilis	Sense Primer	75	100.00%	0.00%	0.00%	0.00%
Bacteroides fragilis	Antisense Primer	75	100.00%	0.00%	0.00%	0.00%
Bacteroides fragilis	Hydrolysis Probe	75	100.00%	0.00%	0.00%	0.00%

10.3.2. Wet-Test Analytical Reactivity

The analytical reactivity (inclusivity) of the ***MarinaBiolab Pre-Plated Vaginal Women Health Panel PCR Kit*** was demonstrated using a comprehensive panel that represents the temporal, evolutionary, and geographic diversity of each target organism.

Each sample was tested in triplicate with the ***MarinaBiolab Pre-Plated Vaginal Women Health Panel PCR Kit*** at an initial concentration 3-fold higher than the LoD determined for each analyte. In cases where the expected targets were not detected in one or more replicates, concentrations 3-fold higher were evaluated.

The individual strains and the concentrations at which positive test results were obtained for all three replicates are presented by target organisms in Table 10 below.

Table 10. Results of the wet inclusivity test.

Variant/Type/Subtype/Lineage/Genotype/Species	Isolate ID/Source	xLoD Detected
Haemophilus ducreyi	Zeptometrix 0801736DNA	1x
Bacterial Vaginosis Associated Bacteria 2	ATCC TSD-275	1x
Fannyhessea vaginae	ATCC BAA-55	1x
Candida albicans	ATCC 10231	1x
Candida glabrata	ATCC 90030	1x
Candida parapsilosis	ATCC 22019	1x
Escherichia coli	ATCC 25922	1x
Streptococcus agalactiae	ATCC 12386	1x
Prevotella bivia	Zeptometrix 0801756	1x
Lactobacillus gasseri	Zeptometrix 0804327	1x
Lactobacillus iners	Zeptometrix 0804261	1x
Lactobacillus crispatus	Zeptometrix 0804143	1x
Megasphaera phylotypes 1	In-house	1x
Megasphaera phylotypes 2	In-house	1x
Staphylococcus aureus	ATCC 12600	1x
Mobiluncus curtisii	Zeptometrix 0804141	1x
Mobiluncus mulieris	Zeptometrix 0804116	1x

Gardnerella vaginalis	ATCC 49145	1x
Enterococcus faecalis	Zeptometrix 0804216	1x
Enterococcus faecium	ATCC BAA-2127	1x
Lactobacillus jensenii	Zeptometrix 0804260	1x
Candida tropicalis	ATCC 750	1x
Trichomonas vaginalis	ATCC 30001	1x
Candida lusitaniae	Zeptometrix 0801603	1x
Herpes Simplex Virus 1	ATCC VR-1778	1x
Herpes Simplex Virus 2	Zeptometrix 0810217CF	1x
Mycoplasma genitalium	ATCC 33530D	1x
Candida dubliniensis	Zeptometrix 0801915	1x
Neisseria gonorrhoeae	ATCC 19424	1x
Candida krusei	ATCC 2159	1x
Chlamydia trachomatis	Zeptometrix 0801775	1x
Treponema pallidum	In-house	1x
Ureaplasma urealyticum	ATCC 27618	1x
Mycoplasma hominis	ATCC 27545-TTR	1x
Bacteroides fragilis	Zeptometrix 0801583	1x

10.4. Analytical Specificity (Exclusivity)

10.4.1. In-Slico Analytical Specificity

Primers and probes designed for a target sequence may also bind to similar sequences if they closely match or differ by only a few base pairs from a non-targeted sequence. To ensure specificity to the target sequence, it is essential to screen the primers and probes against the reference database for the intended templates, as well as any databases that may contain potential contaminating templates.

Table 11. The results of On-Panel and Off-Panel organisms tested for cross-reactivity.

On-Panel/Off-Panel	Name of the organism	Cross Reactivity*		
		Forward	Probe	Reverse
On-Panel	*Haemophilus ducreyi*	None	None	None
On-Panel	*Bacterial Vaginosis Associated Bacteria 2*	None	None	None
On-Panel	*Fannyhessea vaginae*	None	None	None
On-Panel	*Candida albicans*	None	None	None

On-Panel	*Candida glabrata*	None	None	None
On-Panel	*Candida parapsilosis*	None	None	None
On-Panel	*Escherichia coli*	None	None	None
On-Panel	*Streptococcus agalactiae*	None	None	None
On-Panel	*Prevotella bivia*	None	None	None
On-Panel	*Lactobacillus gasseri*	None	None	None
On-Panel	*Lactobacillus iners*	None	None	None
On-Panel	*Lactobacillus crispatus*	None	None	None
On-Panel	*Megasphaera phylotypes 1*	None	None	None
On-Panel	*Megasphaera phylotypes 2*	None	None	None
On-Panel	*Staphylococcus aureus*	None	None	None
On-Panel	*Mobiluncus curtisii*	None	None	None
On-Panel	*Mobiluncus mulieris*	None	None	None
On-Panel	*Gardnerella vaginalis*	None	None	None
On-Panel	*Enterococcus faecalis*	None	None	None
On-Panel	*Enterococcus faecium*	None	None	None
On-Panel	*Lactobacillus jensenii*	None	None	None
On-Panel	*Candida tropicalis*	None	None	None
On-Panel	*Trichomonas vaginalis*	None	None	None
On-Panel	*Candida lusitaniae*	None	None	None
On-Panel	Herpes Simplex Virus 1	None	None	None
On-Panel	Herpes Simplex Virus 2	None	None	None
On-Panel	*Mycoplasma genitalium*	None	None	None
On-Panel	*Candida dubliniensis*	None	None	None
On-Panel	*Neisseria gonorrhoeae*	None	None	None
On-Panel	*Candida krusei*	None	None	None
On-Panel	*Chlamydia trachomatis*	None	None	None
On-Panel	*Treponema pallidum*	None	None	None
On-Panel	*Ureaplasma urealyticum*	None	None	None
On-Panel	*Mycoplasma hominis*	None	None	None
On-Panel	*Bacteroides fragilis*	None	None	None
Off-Panel	*Acinetobacter calcoaceticus*	None	None	None

Off-Panel	*Acinetobacter baumannii*	None	None	None
Off-Panel	*Serratia marcescens*	None	None	None
Off-Panel	*Klebsiella aerogenes*	None	None	None
Off-Panel	*Klebsiella oxytoca*	None	None	None
Off-Panel	*Staphylococcus saprophyticus*	None	None	None
Off-Panel	*Klebsiella pneumoniae*	None	None	None
Off-Panel	*Proteus mirabilis*	None	None	None
Off-Panel	*Proteus vulgaris*	None	None	None
Off-Panel	*Morganella morganii*	None	None	None
Off-Panel	*Citrobacter freundii*	None	None	None
Off-Panel	*Aerococcus urinae*	None	None	None
Off-Panel	*Neisseria meningitidis*	None	None	None
Off-Panel	Human papillomavirus 16	None	None	None
Off-Panel	Human papillomavirus 18	None	None	None
Off-Panel	Human papillomavirus type 52	None	None	None
Off-Panel	Human papillomavirus 6	None	None	None
Off-Panel	Human papillomavirus 11	None	None	None
Off-Panel	Human papillomavirus type 58	None	None	None
Off-Panel	Human papillomavirus type 33	None	None	None

* Homology should be <80% between the cross-reactivity microorganisms and the test primers/ probe(s).

10.4.2. Wet-Test Analytical Specificity

The potential for non-specific amplification by assays designed to detect analytes was evaluated by testing high concentrations of organisms or nucleic acids using the ***MarinaBiolab Pre-Plated Vaginal Women Health Panel PCR Kit***. On-panel organisms were tested to assess potential intra-panel cross-reactivity, while off-panel organisms were tested to evaluate the specificity of the panel. Off-panel organisms included normal flora, pathogens that may be present in specimens, and genetically related species to those detected by the ***MarinaBiolab Pre-Plated Vaginal Women Health Panel PCR Kit***. The concentration of organisms tested (in triplicate) was at least 1.0E+06 CFU/mL for bacteria, fungi, and parasites, and at least 1.0E+05 units/mL for viruses. For certain organisms that were not available for laboratory testing, in silico analysis of the organism's whole genome sequences was used. The on-panel and off-panel organisms tested are listed in Table 12 and Table 13.

Table 12. On-Panel organisms tested for evaluation of ***MarinaBiolab Pre-Plated Vaginal Women Health Panel PCR Kit*** analytical specificity.

Organism	Isolate ID/Source	Cross Reactivity Detected
Haemophilus ducreyi	Zeptometrix 0801736DNA	None
Bacterial Vaginosis Associated Bacteria 2	ATCC TSD-275	None
Fannyhessea vaginae	ATCC BAA-55	None
Candida albicans	ATCC 10231	None
Candida glabrata	ATCC 90030	None
Candida parapsilosis	ATCC 22019	None
Escherichia coli	ATCC 25922	None
Streptococcus agalactiae	ATCC 12386	None
Prevotella bivia	Zeptometrix 0801756	None
Lactobacillus gasseri	Zeptometrix 0804327	None
Lactobacillus iners	Zeptometrix 0804261	None
Lactobacillus crispatus	Zeptometrix 0804143	None
Megasphaera phylotypes 1	In-house	None
Megasphaera phylotypes 2	In-house	None
Staphylococcus aureus	ATCC 12600	None
Mobiluncus curtisii	Zeptometrix 0804141	None
Mobiluncus mulieris	Zeptometrix 0804116	None
Gardnerella vaginalis	ATCC 49145	None
Enterococcus faecalis	Zeptometrix 0804216	None
Enterococcus faecium	ATCC BAA-2127	None
Lactobacillus jensenii	Zeptometrix 0804260	None
Candida tropicalis	ATCC 750	None
Trichomonas vaginalis	ATCC 30001	None
Candida lusitaniae	Zeptometrix 0801603	None
Herpes Simplex Virus 1	ATCC VR-1778	None
Herpes Simplex Virus 2	Zeptometrix 0810217CF	None
Mycoplasma genitalium	ATCC 33530D	None
Candida dubliniensis	Zeptometrix 0801915	None
Neisseria gonorrhoeae	ATCC 19424	None

Candida krusei	ATCC 2159	None
Chlamydia trachomatis	Zeptometrix 0801775	None
Treponema pallidum	In-house	None
Ureaplasma urealyticum	ATCC 27618	None
Mycoplasma hominis	ATCC 27545-TTR	None
Bacteroides fragilis	Zeptometrix 0801583	None

Table 13. Off-Panel organisms were tested for evaluation of ***MarinaBiolab Pre-Plated Vaginal Women Health Panel PCR Kit*** analytical specificity.

Organism	Isolate ID/Source	Cross Reactivity Detected
Acinetobacter calcoaceticus	ATCC 23055	None
Acinetobacter baumannii	ATCC 19606	None
Serratia marcescens	ATCC 29021	None
Klebsiella aerogenes	ATCC 13048	None
Klebsiella oxytoca	ATCC 700324	None
Staphylococcus saprophyticus	Zeptometrix 0804014	None
Klebsiella pneumoniae	NCTC 13465	None
Proteus mirabilis	Zeptometrix 0801544	None
Proteus vulgaris	ATCC 6380	None
Morganella morganii	Zeptometrix 0804010	None
Citrobacter freundii	Zeptometrix 0801563	None
Aerococcus urinae	ATCC 51268	None
Human papillomavirus 16	NIBSC-UK-EN63QG	None
Human papillomavirus 18	NIBSC-UK-EN63QG	None
Human papillomavirus type 52	NIBSC-UK-EN63QG	None
Human papillomavirus 6	NIBSC-UK-EN63QG	None
Human papillomavirus 11	NIBSC-UK-EN63QG	None
Human papillomavirus type 58	NIBSC-UK-EN63QG	None
Human papillomavirus type 33	NIBSC-UK-EN63QG	None
Neisseria meningitidis	ATCC 13090	None

10.5. Interferences

The potential for endogenous or exogenous substances, which may be present in research samples or introduced during sample collection and handling, to interfere with the accurate detection of analytes was evaluated through select direct testing on the ***MarinaBiolab Pre-Plated Vaginal Women Health Panel PCR Kit***. The findings were extrapolated from the interference evaluation of the kit.

Potentially interfering substances were evaluated using contrived samples spiked with the substance of interest. Results from samples containing the substance were compared to those from control samples without the substance. The substances tested included endogenous compounds that may be present in samples at normal or elevated levels (e.g., blood, mucus/mucin, human genomic DNA), various commensal or infectious microorganisms, medications, washes or topical applications, swabs and transport media used for sample collection, and substances employed to clean, decontaminate, or disinfect work areas. Each substance was added to contrived samples containing representative organisms at concentrations near (3x) the LoD. The concentration of each substance added to the samples was equal to or greater than the highest level expected in research samples, and each sample was tested in triplicate.

None of the substances tested were found to interfere with the ***MarinaBiolab Pre-Plated Vaginal Women Health Panel PCR Kit***.

Table 14. Evaluation of potentially interfering substances on the ***MarinaBiolab Pre-Plated Vaginal Women Health Panel PCR Kit***.

Substance Tested	Concentration Tested	Observed Interference
Endogenous Substances		
Human Blood	10% v/v	No Interference
Human Mucus	1 swab/mL sample	No Interference
Human Genomic DNA	20 ng/µL	No Interference
Competitive Microorganisms		
Haemophilus ducreyi	1.0E+06 CFU/mL	No Interference
Bacterial Vaginosis Associated Bacteria 2	1.0E+06 CFU/mL	No Interference
Fannyhessea vaginae	1.0E+06 CFU/mL	No Interference
Candida albicans	1.0E+06 CFU/mL	No Interference
Candida glabrata	1.0E+06 CFU/mL	No Interference
Candida parapsilosis	1.0E+06 CFU/mL	No Interference
Escherichia coli	1.0E+06 CFU/mL	No Interference
Streptococcus agalactiae	1.0E+06 CFU/mL	No Interference
Prevotella bivia	1.0E+06 CFU/mL	No Interference
Lactobacillus gasseri	1.0E+06 CFU/mL	No Interference
Lactobacillus iners	1.0E+06 CFU/mL	No Interference
Lactobacillus crispatus	1.0E+06 CFU/mL	No Interference
Megasphaera phylotypes 1	1.0E+06 CFU/mL	No Interference

Megasphaera phylotypes 2	1.0E+06 CFU/mL	No Interference
Staphylococcus aureus	1.0E+06 CFU/mL	No Interference
Mobiluncus curtisii	1.0E+06 CFU/mL	No Interference
Mobiluncus mulieris	1.0E+06 CFU/mL	No Interference
Gardnerella vaginalis	1.0E+06 CFU/mL	No Interference
Enterococcus faecalis	1.0E+06 CFU/mL	No Interference
Lactobacillus jensenii	1.0E+06 CFU/mL	No Interference
Candida tropicalis	1.0E+06 CFU/mL	No Interference
Trichomonas vaginalis	1.0E+06 CFU/mL	No Interference
Candida lusitaniae	1.0E+06 CFU/mL	No Interference
Herpes Simplex Virus 1	1.0E+05 CFU/mL	No Interference
Herpes Simplex Virus 2	1.0E+05 CFU/mL	No Interference
Mycoplasma genitalium	1.0E+06 CFU/mL	No Interference
Candida dubliniensis	1.0E+06 CFU/mL	No Interference
Neisseria gonorrhoeae	1.0E+06 CFU/mL	No Interference
Candida krusei	1.0E+06 CFU/mL	No Interference
Chlamydia trachomatis	1.0E+06 CFU/mL	No Interference
Treponema pallidum	1.0E+06 CFU/mL	No Interference
Ureaplasma urealyticum	1.0E+06 CFU/mL	No Interference
Mycoplasma hominis	1.0E+06 CFU/mL	No Interference
Bacteroides fragilis	1.0E+06 CFU/mL	No Interference
Exogenous Substances		
K-Y Personal Lubricant Jelly	1% v/v	No Interference
Ortho Options Gynol II Extra Strength Vaginal Contraceptive Jelly	1% v/v	No Interference
Azithromycin	1.8 mg/mL	No Interference
Vagisil Creme Maximum Strength	1% w/v	No Interference
Aspirin	40 mg/mL	No Interference
K-Y Personal Lubricant Jelly	1% v/v	No Interference
Specimen Collection Materials		
Copan Liquid Amies Elution Swab (ESwab®)	N/A	No Interference

11. TROUBLESHOOTING

Problem	Cause	Solution
Target-specific and/or internal control (IC) signals were detected in the Negative Control well.	Contamination may arise from the environment, contamination of extraction and/or qPCR reagents, or well-to-well cross-contamination. The signal observed is not true target amplification, but rather background curves generated by the software of the qPCR instrument.	Repeat the qPCR using fresh reagents. Follow the general GLP guidelines in a PCR lab (e.g., decontaminate all surfaces and instruments with sodium hypochlorite or ethanol, and ensure filter tips are used and changed between samples). It is recommended to set up the qPCR reactions in a separate area, where no RNA/DNA is handled, and with equipment designated solely for pre-PCR activities. Ignore the Cq value of the No Template Control (NTC) if the amplification curve appears to be background noise rather than a true signal. If the issue persists, contact Technical Support.
No IC signal is detected, but a target-specific signal is observed in the sample wells.	A high copy number of target nucleic acid in the samples leads to preferential amplification of the target-specific nucleic acid.	No action is required. The result is considered positive.
The Positive Control did not meet the criteria for acceptable values specified by the kit, rendering the assay invalid.	The Positive Control was not stored under the recommended conditions. The kit has expired.	Check the kit label for the recommended storage conditions and expiration date. Replace the Positive Control. If necessary, use a new kit.
High Cq values were observed in the repeated samples.	The frozen samples were not mixed properly after thawing. Nucleic acids may be degraded.	Ensure frozen samples are thawed with mild agitation to guarantee thorough mixing. Make sure samples are stored correctly and are not subjected to multiple freeze-thaw cycles.
Target-specific and/or IC signals were detected after 35 cycles in the Positive Control.	Incorrect qPCR set-up or the kit reagents may have been compromised (e.g., improper storage or more than 15 freeze-thaw cycles).	Replace the control. If the problem persists, contact Technical Support.
No target-specific or IC signals were detected in the sample wells.	Sampling, extraction, or inhibition problem.	Dilute the nucleic acid isolate 1:10 and repeat the qPCR. If the diluted sample does not show a positive result in the IC channel, request a new sample and repeat the nucleic acid extraction. If necessary, repeat the nucleic acid extraction and the qPCR. If the issue persists, request a new sample, repeat the nucleic acid extraction and qPCR. If the problem continues, contact Technical Support.

12. EXPLANATION of SYMBOLS

Symbol	Title of Symbol	Symbol	Title of Symbol
RUO	Research Use Only		Use-by date
	Manufacturer	LOT	Batch code
CONTROL –	Negative control	NON STERILE	Non-sterile
CONTROL +	Positive control		Consult instructions for use or consult electronic instructions for use
CONTROL	Control		Caution
	Temperature limit	REF	Catalogue number
	Keep away from sunlight		Do not use if package is damaged and consult instructions for use
	Keep dry		Keep upright
Σ	Contains sufficient for <n> tests		Protect from heat and radioactive sources

Custom care and technical support

Tel: +1 510 579-5802

e-mail customer care: accounting@marinabiolab.com

e-mail Technical Support: rd@marinabiolab.com

MarinaBiolab LLC.

Address: 715 Discovery Blvd, suite 309 Cedar Park, TX 78613

For research use only (RUO)! Not for use in diagnostic procedures.