RUO

INSTRUCTION FOR USE

Respiratory Mini Panel PCR Kit

For Research Use Only

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CONTENTS

1. INTENDED USE

For Research Use Only (RUO). Not for use in diagnostic procedures. No claim or representation is intended to provide information for the diagnosis, prevention, or treatment of disease. Furthermore, the test kit is not intended to diagnose infectious animal diseases.

The ***MarinaBiolab Respiratory Mini Panel PCR Kit*** is a multiplexed qualitative Real-Time Reverse Transcription Polymerase Chain Reaction (RT-qPCR) test intended for the simultaneous detection and identification of multiple pathogenic nucleic acids in research samples. The ***MarinaBiolab Respiratory Mini Panel PCR Kit*** allows to achieve RT-qPCR result in less than 1 hour. The test is performed to detect gene sequences of the following organisms.

Targets	Controls
SARS-CoV-2	Human RNase P (IC)
Influenza A	MS2 Bacteriophage (EC)
Influenza B	
H5N1	
Respiratory Syncytial Virus A	
Respiratory Syncytial Virus B	

2. PRINCIPLE of the PROCEDURE

From the RNA and DNA target regions in the lysed or extracted research samples, the RNA target regions are reverse transcribed into cDNA via reverse transcriptase. Then cDNA and DNA target regions are amplified via real-time PCR instruments using the primer and probe sets in the kit. In the process, the probe anneals a specific target sequence located between the forward and reverse primers. During the extension phase of the PCR cycle, the 5' nuclease activity of Taq polymerase degrades the probe, causing the reporter dye to separate from the quencher, generating a fluorescent signal. With each cycle, additional reporter dye molecules are cleaved from their respective probes, increasing the fluorescence intensity. Fluorescence intensity is monitored at each PCR cycle by the real-time PCR instruments. Probes labeled with different fluorophores are used to detect specific amplicons originating from targets and Internal Control.

PCR instruments measure these signals at the end of each amplification cycle in real time and interpret the data to provide a qualitative result for each of the above targets. A positive result for the detection of target RNA or DNA is indicated by the presence of a real-time PCR growth curve and an associated Cq (Quantification Cycle) value.

3. KIT COMPONENTS

The ***MarinaBiolab Respiratory Mini Panel PCR Kit*** consists of four main components:

1. RT-qPCR Enzyme and Buffer Mix (RT-qPCR Master Mix)
2. Forward, Reverse and Probe Oligo Mix (MRPP Oligo Mix 1-2)
3. Mix of non-infectious cDNA and DNA from artificial sample including targets in the table below (PC-MRPP)
4. DNase/RNase-Free Water (NTC)

The kit components are provided in Table 1-2.

Table 1. Kit components.

Component	Description	Quantity x Volume
		100 rxn **MBLRPP002**
RT-qPCR Master Mix	Ready-to-use mix for RT-qPCR	1 x 1000 µL
MRPP Oligo Mix 1-2	Primers and probes complementary to specific regions of the targets in the table above	2 x 250 µL
PC-MRPP	Mix of non-infectious cDNA and DNA from artificial sample including targets in the table above	1 x 400 µL
NTC	DNase/RNase-Free Water	1 x 400 µL

Table 2. Oligo Mix target organisms and detection channels.

Vial Name	Target	Channel
MRPP Oligo Mix 1	SARS-CoV-2	**FAM/Green**
	Influenza A	**HEX/VIC/JOE/Yellow**
	Influenza B	**ROX/Texas Red/Orange**
	MS2 Bacteriophage (EC)	**CY5/Red**
MRPP Oligo Mix 2	H5N1	**FAM/Green**
	Human RNase P (IC)	**HEX/VIC/JOE/Yellow**
	Respiratory Syncytial Virus A	**ROX/Texas Red/Orange**
	Respiratory Syncytial Virus B	**CY5/Red**

The oligonucleotide set targeting the human *RNase P* mRNA (Internal Control: IC) and MS2 Bacteriophage (EC) are used to monitor sampling, nucleic acid extraction, reverse transcription, and inhibition of both reverse transcription and qPCR. The kit also contains negative and positive control templates for evaluating the contamination and the RT-qPCR reagent stability, respectively.

4. EQUIPMENT and MATERIALS REQUIRED but NOT PROVIDED

- 2-8°C Refrigerator
- ≤ -20°C Freezer
- ≤ -70°C Freezer (Optional)
- Vortex mixer
- Benchtop centrifuge with rotor for 1.5 mL tubes
- Benchtop mini centrifuge with rotor for PCR strips
- Benchtop plate centrifuge
- Biological Safety Cabinet (BSC)
- PCR cabinet for PCR Setup
- Adjustable Micropipettes: 1-10, 10-100, 100-1000 µL
- Sterile DNase/RNase free micropipettes tips - Compatible with the micropipettes
- Cold tube rack for microfuge tubes (1.5/2 mL) and for PCR tubes (0.1/0.2 mL)
- Disposable, powder-free, nitrile gloves
- Disposable (preferably) laboratory coat
- Surface decontaminants - Freshly diluted 10% bleach solution (0.5% NaClO)
- Applied Biosystems QuantStudio 5, 7, and 12K with Design & Analysis software and consumables
- Bio-Rad CFX96 Touch™/CFX96™ Dx/CFX Opus 96™/CFX Opus 96™ Dx/CFX384 Touch™/CFX Opus 384™ with Maestro software v1.1 and consumables
- Qiagen Rotor-Gene Q 5plex Platform with Rotor-Gene Q series software v2.1.0.9 and consumables
- Roche LightCycler 480 with software and consumables

5. WARNING and PRECAUTIONS

- The ***MarinaBiolab Respiratory Mini Panel PCR Kit*** is designed for research use only and should be used by professionally trained, qualified staff only. All work should be performed using Good Laboratory Practices.
- Biological material used for extraction of nucleic acid should be handled as potentially infectious. When handling biological material appropriate safety precautions are recommended (do not pipet by mouth; wear disposable gloves while handling biological material and performing the test; disinfect hands when finished the test).
- Biological material should be inactivated before disposal (e.g., in an autoclave). Disposables should be autoclaved or incinerated after use.
- Spillage of potentially infectious materials should be removed immediately with absorbent paper tissue and the contaminated areas swabbed with a suitable standard disinfectant or 70% alcohol. Material used to clean spills, including gloves, should be inactivated before disposal (e.g., in an autoclave).
- Disposal of all samples, unused reagents and waste should be in accordance with country, federal, state, and local regulations.
- Microbial contamination of the reagents while taking aliquots should be avoided. It is recommended to use sterile one-way pipettes and tips. Reagents that look cloudy or show any signs of microbial contamination must not be used.
- The kit should be stored away from nucleic acid sources and qPCR amplicons.
- Always check the expiration date on the kit. Do not use expired or incorrectly stored kit.
- The components in the kit should not be mixed with components with different lot numbers or chemicals of the same name but from different manufacturers.
- Kit components should be mixed by gently shaking before use.
- A common concern with PCR-based assays is false positive results caused by contamination of the work area with PCR amplicon. To prevent amplicon contamination:
    - It shall be ensured that separate work areas with their own apparatus are available.
    - Do not open the reaction tubes/plates post amplification to avoid contamination with amplicons.
    - Discard used tubes/plates in a biohazard container immediately after the run has completed.
    - Avoid excessive handling of tubes/plates after test runs.
    - Change gloves after handling a used tubes/plate.

6. HANDLING, STORAGE, and STABILITY

- The ***MarinaBiolab Respiratory Mini Panel PCR Kit*** is shipped on dry ice. If any component except RT-qPCR Master Mix of the kit is not frozen on arrival, or if the outer packaging has been compromised during shipment, please contact **MarinaBiolab** or the local distributors as soon as possible.
- All components should be stored between -25°C and -15°C upon arrival.
- Repeated freezing and thawing of the kit components may result in lower detection quality. The kit can undergo up to 15 freeze/thaw cycles without affecting performance.
- When stored under the specified storage conditions, the kit is stable until the stated expiration date printed on the package. The expiration date of the kit is 12 months from date of manufacture.
- All components must be thawed at ambient temperature for a minimum of 30 minutes before use.
- It is recommended that all components should be kept on ice when setting up the assay mixes.
- The primer and probe mixes contain fluorophore labeled probes and should be protected from direct sunlight or long-term ambient light.
- Do not use expired or incorrectly stored components.

7. TEST PROCEDURE

7.1. Sample Preparation and Nucleic Acid Extraction

The sample material for the isolation of nucleic acid must be sent in appropriate cell collection systems. The performance of the kit strongly depends on the amount and quality of the extracted nucleic acid. It must be ensured that the system used for nucleic acid extraction is compatible with real-time PCR technology.

If the established standard method of the lab is used for nucleic acid isolation, it must be validated by the user.

For frozen samples or frozen extracted nucleic acid, only thaw the number of specimen extracts that will be tested in a single day.

Do not freeze/thaw extracted nucleic acid more than once before testing as each freeze/thaw cycle can decrease the nucleic acid quality. For optimal results, use it directly.

7.2. PCR Reaction Preparation and Processing

- Completely thaw the components at room temperature for a minimum of 30 minutes before each use.
- Place all components on ice once thawed during the whole test procedure.
- Determine the number of reactions and create the PCR plate plan.
- Include the following reactions to the plan:
    - Reactions for each test sample and extraction negative control.
    - PCR control reactions:
        - Positive Control (included in the kit)
        - Negative (No Template) Control (NTC) (included in the kit)
        - No Template Addition Control (NRC)
- Vortex and spin down briefly the components before each use.
- Combine the following components for the number of reactions required plus 10% overage to compensate for pipetting errors:

Table 3. Reaction set-up.

Reaction Mix Component	1X Reaction (µL) per well
RT-qPCR Master Mix	5 µL
MRPP Oligo Mix 1-2	2.5 µL
Template Nucleic Acid	2.5 µL
Total Reaction Volume	10 µL

- Add 5 µL of RT-qPCR Master Mix and 2.5 µL of MRPP Oligo Mix 1-2 into PCR tubes.
- Add 2.5 µL of the isolated sample into the individual tubes.
- The final reaction mix volume is 10 µL.
- Close the tubes, centrifuge briefly, insert tubes into the real-time PCR instrument and amplify according to the following PCR profile.

Table 4. Amplification profile.

Step	Number of Cycles	Temperature	Time	Data Collection
Reverse Transcription	1	52 °C	5 min	**FAM/Green**, **HEX/VIC/JOE/Yellow**, **ROX/Texas Red/Orange**, **CY5/Red**
Initial Denaturation	1	95 °C	10 sec	
Denaturation	40	95 °C	5 sec	
Annealing/Extension		55 °C	15 sec	

8. INTERPRETATION OF RESULTS

MarinaBiolab Respiratory Mini Panel PCR Kit provides a qualitative result for the presence (Detected) or absence (Not Detected) of the target genes.

8.1. Calculation of Cq Values and Instrument-Specific Requirements

Perform the following instrument settings before evaluating the results.

Table 5. Instrument-specific requirements before evaluating the results.

Instrument	Threshold Level	Other Settings
CFX96 Touch™/CFX96™ Dx/CFX Opus 96™/CFX Opus 96™ Dx/ CFX384 Touch™/CFX Opus 384™ (Bio-Rad)	500 RFU	-
Rotor-Gene Q 5plex Platform (QIAGEN)	0.02 RFU	Dynamic Tube: Active Slope Correct: Active Outlier Removal: 0
QuantStudio™ 5, 7 and 12K (Applied Biosystems™)	Auto	-
Roche LightCycler 480 (Roche)	Auto	-

The shape of the amplification curves should be examined. If a Cq value is assigned to a sample by the instruments' software and the curve is sigmoidal, the Cq value can be used in the final evaluation. *Non-sigmoidal curves should be recorded as negative.*

The result is recorded as positive if Cq≤38 or as established by your lab.

8.2. Overall Validity of Detection

Table 6. Expected performance of controls.

Control Type	Used to Monitor	Signal	
		Target Channel	Internal/External Control Channel
Negative Control	Cross-contamination during extraction and reaction setup	-	-
No template addition	Reagent and/or environmental contamination	-	-
Positive Control	RT-qPCR reaction setup and reagent integrity	+	+
Internal/External Control	To monitor the integrity of nucleic acid extraction and RT-qPCR from each specimen	Not applicable	+

Before analyzing samples results, we recommend verifying if the real-time PCR test is valid. Thus, for each run, please confirm if the results for Positive and Negative controls performed as expected, according to the following criteria:

Table 7. Run validity/positive and negative control pass criteria.

Positive Control		Negative Control		Results	Recommendation
Target Channel	**Internal/External Control Channel**	**Target Channel**	**Internal/External Control Channel**		
+	+	-	-	VALID	Continue to result interpretation of samples.
Any of them is Negative		Not considered		INVALID	Contact the manufacturer, renew the reagents, and repeat the reaction.
Not considered		Any of them is Positive		INVALID	Repeat analysis, paying attention to "Warnings and Precautions" in IFU.

If any control does not perform as described above, the run is considered invalid, and the test is repeated. If the problem persists, contact the manufacturer.

If all the controls are valid, proceed to the interpretation of the results.

8.3. Interpretation of Unknown Specimen Results

The data produced by the instruments can manually be evaluated and reported using their software.

Table 8. Interpretation of unknown specimen results for pathogens.

RNA Pathogens	Internal Control (*RNase P*)	External Control (*MS2*)	Results	Interpretation
Positive (+) (Cq<38)	Positive (+) (Cq<38)	Positive (+) (Cq<38)	Positive for Target	Target RNA is detected
Positive (+) (Cq<38)	Negative (-) (Cq≥38 or N/A)	Positive (+) (Cq<38)	Positive for Target	Target RNA is detected
Positive (+) (Cq<38)	Negative (-) (Cq≥38 or N/A)	Negative (-) (Cq≥38 or N/A)	Invalid	Repeat test by re-extracting the sample. If the repeat result remains invalid, consider collecting a new sample.
Positive (+) (Cq<38)	Positive (+) (Cq<38)	Negative (-) (Cq≥38 or N/A)	Invalid	Repeat test by re-extracting the sample. If the repeat result remains invalid, consider collecting a new sample.
Negative (-) (Cq≥38 or N/A)	Positive (+) (Cq<38)	Positive (+) (Cq<38)	Negative for Target	Target RNA is not detected
Negative (-) (Cq≥38 or N/A)	Negative (-) (Cq≥38 or N/A)	Positive (+) (Cq<38)	Negative for Target	Target RNA is not detected
Negative (-) (Cq≥38 or N/A)	Negative (-) (Cq≥38 or N/A)	Negative (-) (Cq≥38 or N/A)	Invalid	Repeat test by re-extracting the sample. If the repeat result remains invalid, consider collecting a new sample.
Negative (-) (Cq≥38 or N/A)	Positive (+) (Cq<38)	Negative (-) (Cq≥38 or N/A)	Invalid	Repeat test by re-extracting the sample. If the repeat result remains invalid, consider collecting a new sample.

9. ASSAY LIMITATIONS

- The ***MarinaBiolab Respiratory Mini Panel PCR Kit*** is intended for use by professionally trained, qualified staff only.
- A false negative result may occur if a specimen is improperly collected, transported, or handled. False negative results may also occur if amplification inhibitors are present in the specimen or if inadequate numbers of organisms are present in the specimen.
- Spontaneous mutations within the target sequences may result in failure to detect the target sequence. While this risk is mitigated in the test's design, if failure to detect the target is expected it is recommended to test the specimen with a different test that detects different target sequences from the target's genome.
- There is a risk of false positive results due to cross-contamination by target viruses and/or bacteria, their nucleic acids or amplified product, or from non-specific signals in the assay. Attention should be given to the handling of consumables under the Warnings and Precautions section to help minimize this risk.
- This assay is a qualitative test and does not provide a quantitative assessment of the concentration of the detected organism.
- All instruments (e.g., pipettes, real-time cyclers) must be calibrated according to the manufacturer's instructions.

10. PERFORMANCE CHARACTERISTICS

10.1. Analytical Sensitivity (Limit of Detection, LoD)

The LoD was defined as the concentration at which the test produces a positive result >95% of the time. Serial dilutions of the strains were tested and the initial tentative LoD confirmed with twenty (20) replicates. To ensure the accuracy of the LoD determination, if the initial detection rate was 100%, a further twenty (20) replicates were performed at the next lower concentration until ≤95% was achieved. For nucleic acid extraction, simulated research matrix was spiked with strains and loaded onto the Automatic Nucleic Acids Extraction Instrument. The tests were carried out using the CFX96 Touch™ (Bio-Rad) Real-Time PCR system. The confirmed LoDs for the strains tested and the corresponding LoDs for the ***MarinaBiolab Respiratory Mini Panel PCR Kit*** reportable targets are shown in Table 9 below.

Table 9. Summary of LoD study results.

Analyte	Isolate ID/Source	LoD Concentration (copies/mL)	Detected/Total
SARS-CoV-2	ATCC VR-1986HK	1.2E+02 copies/mL	**20/20** 100%
Influenza A	Zeptometrix 0810036CF	1.1E+02 copies/mL	**20/20** 100%
Influenza B	Zeptometrix 0810255CF	8.7E+01 copies/mL	**20/20** 100%
Respiratory Syncytial Virus A	Zeptometrix 0810040ACF	9.9E+01 copies/mL	**20/20** 100%
Respiratory Syncytial Virus B	Zeptometrix 0810040CF	1.3E+02 copies/mL	**20/20** 100%
H5N1	Vircell MBC052	7.8E+01 copies/mL	**20/20** 100%

10.2. Device Equivalence Study

Device equivalence study was carried out to observe the differences between the results to be obtained using the kit in different instruments. For this purpose, the same LoD determination study was performed again with the Bio-Rad CFX96™ Dx/CFX Opus 96™/CFX Opus 96™ Dx/CFX384 Touch™/CFX Opus 384™, Applied Biosystems QuantStudio 5, 7, and 12K, Qiagen Rotor-Gene Q 5plex Platform and Roche LightCycler 480. Similar test results were obtained with the 1x LoD concentration level of the targets in the "device equivalence study" performed with the other instruments.

10.3. Analytical Reactivity (Inclusivity)

10.3.1. In-Slico Analytical Reactivity

BLAST search of the oligonucleotides was performed on the SARS-CoV-2, Influenza A, Influenza B, Respiratory Syncytial Virus A, Respiratory Syncytial Virus B, and H5N1 genome sequences available in the NCBI database, using the Primer-BLAST tool of NCBI.

The aggregated result of all in-silico analyzes performed in NCBI database is provided in Table below. The melting temperatures (Tm) of the oligonucleotide sequences with 1-base mismatch, are still higher than the annealing temperature specified in the PCR cycle parameters of the kit. Hence, the single mismatches in the sequences are not expected to affect the inclusivity of the test.

Table 10. In-silico analysis results performed in the NCBI database.

Target	Primer	Total number of target sequences	Ratio of the sequences without mismatch	Ratio of the sequences with 1 base mismatch	Ratio of the sequences with 2 base mismatches	Ratio of the sequences with 3 base mismatches
SARS-CoV-2	ORF1ab Sense Primer	54254	96.60%	3.40%	0.00%	0.00%
SARS-CoV-2	ORF1ab Antisense Primer	54254	96.42%	3.56%	0.02%	0.00%
SARS-CoV-2	ORF1ab Hydrolysis Probe	55425	95.62%	4.38%	0.00%	0.00%
SARS-CoV-2	N Sense Primer	54626	96.42%	3.58%	0.00%	0.00%
SARS-CoV-2	N Antisense Primer	54626	97.27%	2.63%	0.00%	0.00%
SARS-CoV-2	N Hydrolysis Probe	54988	95.45%	4.55%	0.00%	0.00%
Influenza A	Sense Primer	47.865	92.24%	7.65%	0.11%	0.00%
Influenza A	Antisense Primer	47.865	97.52%	2.48%	0.00%	0.00%
Influenza A	Hydrolysis Probe	49.224	96.56%	3.44%	0.00%	0.00%
Influenza B	Sense Primer	7.945	98.89%	1.11%	0.00%	0.00%
Influenza B	Antisense Primer	7.945	99.87%	0.13%	0.00%	0.00%
Influenza B	Hydrolysis Probe	7.947	99.57%	0.43%	0.00%	0.00%
Respiratory Syncytial Virus A	Sense Primer	4615	98.42%	1.58%	0.00%	0.00%
Respiratory Syncytial Virus A	Antisense Primer	4615	98.42%	1.58%	0.00%	0.00%
Respiratory Syncytial Virus A	Hydrolysis Probe	4618	97.46%	2.54%	0.00%	0.00%
Respiratory Syncytial Virus B	Sense Primer	8314	98.66%	1.34%	0.00%	0.00%
Respiratory Syncytial Virus B	Antisense Primer	8314	97.76%	2.24%	0.00%	0.00%
Respiratory Syncytial Virus B	Hydrolysis Probe	8509	98.12%	1.88%	0.00%	0.00%
Human Metapneumovirus A	Sense Primer	1502	97.56%	2.44%	0.00%	0.00%
Human Metapneumovirus A	Antisense Primer	1502	99.81%	0.19%	0.00%	0.00%
Human Metapneumovirus A	Hydrolysis Probe	1504	99.85%	0.15%	0.00%	0.00%
Human Metapneumovirus B	Sense Primer	1341	97.55%	2.45%	0.00%	0.00%
Human Metapneumovirus B	Antisense Primer	1341	97.55%	2.45%	0.00%	0.00%
Human Metapneumovirus B	Hydrolysis Probe	1104	99.28%	0.72%	0.00%	0.00%
H5N1	Sense Primer	7521	97.26%	2.20%	0.54%	0.00%
H5N1	Antisense Primer	7521	97.26%	2.20%	0.54%	0.00%

H5N1	Hydrolysis Probe	7489	97.12%	2.00%	0.88%	0.00%

10.3.2. Wet-Test Analytical Reactivity

The analytical reactivity (inclusivity) of the ***MarinaBiolab Respiratory Mini Panel PCR Kit*** was demonstrated with a comprehensive panel representing temporal, evolutionary, and geographic diversity for each of the target organisms.

Each sample was tested with the ***MarinaBiolab Respiratory Mini Panel PCR Kit*** in triplicate at an initial concentration 3-fold higher than the LoD determined for each analyte. In cases where the expected targets were not detected in one or more replicates, concentrations at a 3-fold higher level were evaluated.

The individual strains and concentrations at which positive test results were obtained for all three (3) replicates are presented by target organism in Table 11 below.

Table 11. Results of the wet inclusivity test.

Variant/Type/Subtype/Lineage/Genotype/Species	Isolate ID/Source	xLoD Detected
SARS-CoV-2	ATCC VR-1986HK	1x
SARS-CoV-2 Delta	ATCC VR-3342HK	1x
SARS-CoV-2 Omicron	ATCC VR-3378HK	1x
Influenza A H1N1	Zeptometrix 0810036CF	1x
Influenza A H1N1pdm09	Zeptometrix 0810109CFJ	1x
Influenza A H3N2	Zeptometrix 0810252CF	3x
Influenza B	Zeptometrix 0810255CF	1x
Influenza B Victoria	Zeptometrix 0810258CF	3x
Influenza B Yamagata	Zeptometrix 0810256CF	3x
Respiratory Syncytial Virus A	Zeptometrix 0810040ACF	1x
Respiratory Syncytial Virus B	Zeptometrix 0810040CF	1x
H5N1	Vircell MBC052	1x

10.4. Analytical Specificity (Exclusivity)

10.4.1. In-Slico Analytical Specificity

Primers and probes intended for a target sequence may also attach to similar sequences if they closely match or differ by only a few base pairs from the non-targeted sequence. To ensure specificity to the target amplicon sequence, it's essential to screen the primers and probe against the reference database transcript or genome database for the intended templates, as well as any databases containing potential contaminating templates.

Table 12. The results of On-Panel and Off-Panel organisms tested for cross-reactivity.

On-Panel/Off-Panel	Name of the organism	Cross Reactivity*		
		Forward	Probe	Reverse
On-Panel	SARS-CoV-2	None	None	None
On-Panel	Influenza A	None	None	None
On-Panel	Influenza B	None	None	None
On-Panel	Respiratory Syncytial Virus A/B	None	None	None
On-Panel	H5N1	None	None	None
Off-Panel	Bat SARS-like Coronavirus	None	None	None
Off-Panel	MERS-CoV	None	None	None
Off-Panel	SARS	None	None	None
Off-Panel	Cytomegalovirus	None	None	None
Off-Panel	Epstein-Barr Virus	None	None	None
Off-Panel	Herpes Simplex Virus 1	None	None	None
Off-Panel	Herpes Simplex Virus 2	None	None	None
Off-Panel	Human Herpes Virus 6	None	None	None
Off-Panel	Measles Virus	None	None	None
Off-Panel	Mumps	None	None	None

* Homology should be <80% between the cross-reactivity microorganisms and the test primers/ probe(s).

** In silico sequence analysis indicates the potential for cross-reactivity of Bordetella pertussis with certain strains of Bordetella bronchiseptica.

10.4.2. Wet-Test Analytical Specificity

The potential for non-specific amplification by assays for detection of analytes was evaluated by testing high concentrations of organisms or nucleic acids with the ***MarinaBiolab Respiratory Mini Panel PCR Kit***. On-panel organisms were tested to assess the potential for intra-panel cross-reactivity, and off-panel organisms were tested to evaluate panel specificity. Off-panel organisms included normal flora and pathogens that may be present in specimens as well as near-neighbors or species genetically related to the organisms detected by the ***MarinaBiolab Respiratory Mini Panel PCR Kit***. The concentration of organism tested (in triplicate) was at least 1.0E+06 CFU/mL for bacteria, fungi and parasite, and at least 1.0E+05 unit/mL for viruses. For the few organisms of interest that were not available for laboratory testing, results of in silico analysis of the organism's whole genome sequences are indicated. The on-panel and off-panel organisms tested are shown in Table 13 and Table 14.

Table 13. On-Panel organisms tested for evaluation of ***MarinaBiolab Respiratory Mini Panel PCR Kit*** analytical specificity.

Organism	Isolate ID/Source	Cross Reactivity Detected
SARS-CoV-2	ATCC VR-1986HK	None
SARS-CoV-2 Delta	ATCC VR-3342HK	None

SARS-CoV-2 Omicron	ATCC VR-3378HK	None
Influenza A H1N1	Zeptometrix 0810036CF	None
Influenza A H1N1pdm09	Zeptometrix 0810109CFJ	None
Influenza A H3N2	Zeptometrix 0810252CF	None
Influenza B	Zeptometrix 0810255CF	None
Influenza B Victoria	Zeptometrix 0810258CF	None
Influenza B Yamagata	Zeptometrix 0810256CF	None
Respiratory Syncytial Virus A	Zeptometrix 0810040ACF	None
Respiratory Syncytial Virus B	Zeptometrix 0810040CF	None
H5N1	Vircell MBC052	None

Table 14. Off-Panel organisms were tested for evaluation of ***MarinaBiolab Respiratory Mini Panel PCR Kit*** analytical specificity.

Organism	Isolate ID/Source	Cross Reactivity Detected
Cytomegalovirus	ATCC VR-977	None
Epstein Barr Virus	Zeptometrix 0810008CF	None
Measles	Zeptometrix 0810025CF	None
Mumps virus	Zeptometrix 0810079CF	None
Varicella-Zoster virus	Zeptometrix 0810026CF	None
Herpes Simplex Virus 1 (HSV1)	ATCC VR-1778	None
Herpes Simplex Virus 2 (HSV2)	Zeptometrix 0810217CF	None

10.5. Interferences

The ability of endogenous or exogenous substances that could be present in research samples (or introduced during sample collection and handling) to interfere with accurate detection of analytes was evaluated with select direct testing on the ***MarinaBiolab Respiratory Mini Panel PCR Kit*** and extrapolated from the interference evaluation of the ***MarinaBiolab Respiratory Mini Panel PCR Kit***.

Potentially interfering substances were evaluated using contrived samples spiked with substance. Results from samples containing a substance were compared to results from control samples without substance. The substances tested included endogenous substances that may be found in sample at normal or elevated levels (e.g., blood, mucus/mucin, human genomic DNA), various commensal or infectious microorganisms, medications, washes or topical applications, various swabs and transport media for sample collection, and substances used to clean, decontaminate, or disinfect work areas. Each substance was added to contrived samples containing representative organisms at concentrations near (3x) LoD. The concentration of substance added to the samples was equal to or greater than the highest level expected to be in research sample and each sample was tested in triplicate.

None of the substances were shown to interfere with the ***MarinaBiolab Respiratory Mini Panel PCR Kit***.

Table 15. Evaluation of potentially interfering substances on the ***MarinaBiolab Respiratory Mini Panel PCR Kit***.

Substance Tested	Concentration Tested	Observed Interference
Endogenous Substances		
Human Blood	10% v/v	No Interference
Human Mucus (Sputum)	1 swab/mL sample	No Interference
Human Genomic DNA	20 ng/µL	No Interference
Competitive Microorganisms		
SARS-CoV-2	1.0E+05 unit/mL	No Interference
Influenza A	1.0E+05 unit/mL	No Interference
Influenza B	1.0E+05 unit/mL	No Interference
Respiratory Syncytial Virus A	1.0E+05 unit/mL	No Interference
Respiratory Syncytial Virus B	1.0E+05 unit/mL	No Interference
H5N1	1.0E+05 unit/mL	No Interference
Exogenous Substances		
Otrivine Adult Nasal Spray	1% v/v	No Interference
Tobramycin (systemic antibiotic)	1 mg/mL	No Interference
Amoxicillin + Penicillin + Cefadroxil + Erythromycin mixture	1% w/v	No Interference
Petroleum Jelly (Vaseline®)	1% w/v	No Interference
Rapivab (peramivir)	1% w/v	No Interference
Specimen Collection Materials		
Nylon Flocked Swabs (Copan 553C)	N/A	No Interference
Calcium Alginate Swabs (Puritan 25-801 A 50)	N/A	No Interference
Polyester Swabs (Copan 175KS01)	N/A	No Interference
Polyester Swabs (Copan 175KS01)	100%	No Interference
Copan ESwab™ Sample Collection and Delivery System (Swab and Liquid Amies Medium)	100%	No Interference

11. TROUBLESHOOTING

Problem	Cause	Solution
Target-specific and/or IC signals are detected in the Negative Control well.	Contamination from the environment, contamination of extraction and/or RT-qPCR reagents, or well-to-well cross contamination. The signal is not true target amplification, but background curves generated by the software of the qPCR instrument.	Repeat the RT-qPCR with new reagents. Follow the general rules of GLP in a PCR lab (e.g., Decontaminate all surfaces and instruments with sodium hypochlorite or ethanol. Ensure that filters tips are used during the procedure and changed between samples.). It is recommended to set up the RT-qPCR reactions in a separate area, where no RNA/DNA is handled and with equipment designated for pre-PCR activities. Ignore the Cq value of NTC if the amplification curve looks not real but background noise. If the problem persists, contact Technical Support.
No IC signal is detected, but target-specific signal is detected in sample wells.	A high copy number of target nucleic acid exists in samples, resulting in preferential amplification of the target-specific nucleic acid.	No action is required. The result is considered positive.
The Positive Control did not meet the criteria set for acceptable values of the kit. The assay is invalid.	Positive Control was not stored at the recommended conditions. Kit shelf-life expired.	Check the kit label for storage conditions and expiration date. Replace the Positive Control. Use a new kit if necessary.
High Cq values observed for repeated samples.	Frozen samples were not mixed properly after thawing. Degraded nucleic acids.	Make sure, thaw frozen samples with mild agitation to ensure thorough mixing. Ensure that samples are stored correctly and not subjected to multiple freeze-thaw cycles
Target-specific and/or IC signal detected after 38 Cycles in Positive Control.	Incorrect RT-qPCR set-up or the kit reagents may have been compromised (e.g., improper storage or more than 15 freeze-thaw cycles).	Replace the control. If the problem persists, contact Technical Support.
No target-specific and IC signal is detected in sample wells.	Sampling, extraction, or inhibition problem.	Dilute the nucleic acid isolate 1/10 and repeat the RT-qPCR. If the diluted sample does not give a positive result in the IC channel, request for a new sample and repeat the NA extraction. Repeat the NA extraction and the RT-qPCR. Request for a new sample, repeat the NA extraction and the RT-qPCR. If the problem persists, contact Technical Support.

12. EXPLANATION of SYMBOLS

Symbol	Title of Symbol	Symbol	Title of Symbol
	Research Use Only		Use-by date
	Manufacturer		Batch code
	Negative control		Non-sterile
	Positive control		Consult instructions for use or consult electronic instructions for use
	Control		Caution
	Temperature limit		Catalogue number
	Keep away from sunlight		Do not use if package is damaged and consult instructions for use
	Keep dry		Keep upright
	Contains sufficient for <n> tests		Protect from heat and radioactive sources

Custom care and technical support

Tel: +1 510 579-5802

e-mail customer care: accounting@marinabiolab.com

e-mail Technical Support: rd@marinabiolab.com

MarinaBiolab LTD.

Address: Hürriyet Mah. Erler So. No: 4/1A Nexus No: 13 Kartal/Istanbul/Turkey

For research use only (RUO)! Not for use in diagnostic procedures.